



## Harnessing heavy metal-tolerant bacteria and phytotoxicity assessment for ecofriendly treatment of industrial effluents

Sharmila Shashikant Ghangale<sup>1\*</sup>, Ramling Sidaramappa Saler<sup>2</sup>, Sharad Ratan Khandelwal<sup>3</sup>, Dilip Vishwanath Handore<sup>4</sup>, Anita Vishwanath Handore<sup>4</sup>

<sup>1</sup> Department of Biotechnology, Changu Kana Thakur Arts, Commerce and Science College, New Panvel, India

<sup>2</sup> Department of Environmental Science, K.T.H.M. College, Nashik, India

<sup>3</sup> Department of Microbiology, Phytoelixir Pvt. Ltd., Nashik, India

<sup>4</sup> Research and Development Department, Phytoelixir Pvt. Ltd., Nashik, India

\* Corresponding author E-mail: gssharmi@yahoo.com

## Article info

Received 4/3/2024; received in revised form 11/4/2024; accepted 30/4/2024 DOI: 10.6092/issn.2281-4485/19182 © 2024 The Authors.

### Abstract

This study treated heavy-metal pollution in industrial wastewater by isolating and characterizing heavy metaltolerant bacterial strains for bioremediation of industrial wastewater. The effluent samples were sterilized and aseptically inoculated with isolated heavy metal-tolerant bacteria such as *Bacillus lichniformis, Pseudomonas aeruginosa, and Stenotrophomonas maltophilia*. The samples were subjected to physicochemical analysis, and titrimetric analysis. Further, Atomic Absorption Spectroscopy (AAS) was used for Heavy metal analysis. Phytotoxic assay was carried out by seed germination tests and pot culture study. Over 96-hour period, improvement featuring reduced turbidity, a stabilized neutral pH, and a substantial 72% decrease in total solids was observed. The application of the bioremediation techniques using heavy metal tolerant bacteria resulted in a significant reduction of heavy metal concentrations, surpassing 85%. Moreover, seed germination rates in the treated industrial effluent were higher than the untreated samples. Pot culture tests further revealed diminished toxicity (2.5%) in treated roots compared to untreated ones (36.7%), accompanied by increased root and shoot growth in the treated pots. The noticeable enhancement in germination rates and the improved growth of plants in the treated effluent underscore the promising potential for the sustainable application of these eco-friendly bacteria, making them a cost-effective microbial bioremediation solution for industrial wastewater.

### Keywords

Eco-friendly, Industrial wastewater, Heavy metals, Microbial Bioremediation, Phytotoxicity.

## **Introduction**

The presence of heavy metals in industrial effluents poses a considerable threat to aquatic ecosystems, human health, and overall ecological balance (Tayang and Songachan, 2021). Conventional wastewater treatment methods often prove inadequate in effectively addressing the intricate composition of industrial effluents. Consequently, there is a pressing need to shift towards ecologically conscious and economically viable alternatives (Ianeva, 2009).

In light of the escalating industrial activities and the consequent release of diverse pollutants into aquatic ecosystems, innovative and sustainable approaches are urgently required to tackle the environmental challenges posed by industrial effluents (Samadi et al., 2015). This study aims to tackle the issue of heavy metal pollution in industrial wastewater by isolating and characterizing heavy metal-tolerant bacterial strains. The primary obje-

ctives include identifying bacterial isolates, enriching them, applying them in the bioremediation process, assessing enzymatic activity, and exploring how variations in pH and temperature influence the efficacy of these heavy metal-tolerant strains in microbial bioremediation.

The study explores the promising realm of microbial bioremediation as a pivotal strategy for mitigating the impact of industrial effluents on water quality (Iwasaki et al., 2009). Focusing on metal-tolerant microbial isolates, the study delves into their potential applications in restoring water ecosystems, offering hope for sustainable water management in eco-friendly way (Ayaz et al., 2020). It not only demonstrates the bioremediation potential of bacterial isolates uniquely adapted to thrive in metal-contaminated environments but also extends its scope to Phytotoxicity assessment(Ji and Silver, 1995). Recognizing the intricate interplay between microbial activity and plant health, the research explores the potential effects of metal-tolerant bacterial isolates on plant physiology (Tolbatov et al., 2020). By evaluating the Phytotoxicity of these isolates, the study aims to provide a holistic understanding of their environmental impact, contributing valuable insights for the development of sustainable water management practices (Hobman and Brown, 1996). As the world grapples with the imperative to safeguard water resources and preserve ecological integrity, the findings presented in this manuscript represent a significant stride towards achieving these objectives (Manisha et al, 2011; Tayang and Songachan, 2021; Yumpu, 2023).

By unravelling the potential of metal-tolerant bacterial isolates in the bioremediation of industrial effluents and concurrently assessing their Phytotoxicity, this research seeks to provide a robust foundation for the development of eco-friendly and efficient wastewater treatment strategies, fostering a harmonious coexistence between industrial progress and environmental sustainnability (Manisha et al, 2011; Tayang and Songachan, 2021; Yumpu, 2023). Therefore, this study emphasizes the promising potential for the sustainable utilization of these eco-friendly bacterial strains for the microbial bioremediation of industrial wastewater.

### Materials and Methods

#### Collection and bioremediation of industrial effluent

Samples were collected in sterile glass bottles from Metallurgical Industries, Nashik. These samples were sterilized and aseptically inoculated with isolated heavy metal-tolerant bacteria i.e., *Bacillus lichniformis, Pseudomonas aeruginosa,* and *Stenotrophomonas maltophilia*. Further, allowed to incubate at 200 rpm. on a rotary shaker at room temperature. Sterilized industrial effluent was kept in control.

#### Physiochemical analysis

The pH, temperature, and electrical conductivity readings were recorded at the sample collection site using respective portable handheld instruments. Total Dissolved Solids (TDS) and Total Suspended Solids (TSS) were determined by gravimetric analysis using the following equations respectively as,

Total Dissolved Solids (TDS) =

Sample volume

[2]

00

Total Suspended Solids (TSDS) =

{[Weight of the Whatman filter paper after filtration (mg)] - [Weight of the Whatman filter paper before filtration (mg)]} \* 100

Sample volume

And Total Solids (mg/L) were calculated by summation of TDS and TSS values.

ł

Thereafter, Dissolved Oxygen (DO) in the industrial effluent sample was assessed using the Winkler's titrimetric method and Biological Oxygen Demand (BOD) was determined using the 5-day incubation method prescribed by the American Public Health Association (APHA) standard. Further, BOD values were computed using the formula: BOD (5d)  $O_2$  mg/L = 16 (V1–V2), where V1 and V2 denote the amounts of Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> required for the sample before and after incubation on the 5th day.

Chemical Oxygen Demand (COD) was determined by the APHA standard titrimetric analysis. COD values were computed using the formula: (A-B)/L =sample (ml) × M × 8000, where A and B represent the amounts of FAS required for the blank and sample, M is the molarity of FAS, and 8000 is the milliequivalent weight of oxygen × 1000 ml/L.

Heavy metal analysis of the industrial effluent sample was performed using Atomic Absorption Spectroscopy (AAS).

# Phytotoxic assay: Seed germination test for treated industrial effluent

Seeds of various dicot and monocot plants viz.maize, green gram, moth beans, cowpea seeds, horse gram, and fenugreek were surface sterilized with 0.1% w/v aqueous solution of Mercuric chloride and rinsed with distilled water. During this test, 10 uniform seeds of each plant were plated on sterilized petri plates with blotting paper, irrigated daily with 5 ml of treated industrial effluent, and incubated at  $25 \pm 2^{\circ}$ C for 3-4 days. Seed germination rates were recorded after every 4 days for treated, untreated, and control groups, providing a systematic evaluation of industrial effluent impact on different plant varieties. Percent germination was calculated as the ratio of germinated seeds by total number of seeds.

## Phytotoxic assay: Pot culture test for treated industrial effluent

Wheat seeds (*Triticum aestivum* L.) were soaked in sterilized distilled water for 5-6 hours and sown individually in six pots filled with clean soil for each treatment group: treated, untreated, and tap water (control). Daily irrigation with 25 ml of treated or untreated industrial effluents was administered to respective pots, while the control pots were irrigated with tap water. Germination rates and seedling growth were monitored daily for 7-12 days. Growth parameters, including shoot length, root length, and shoot height, were measured and recorded for each pot plant.

### Statistical analysis

All experiments were conducted in triplicates. Data was employed for calculating descriptive statistical values. All values are represented as Mean (SD), specified otherwise.

## <u>Results</u>

# Physicochemical characteristics, bioremediation, and heavy metal analysis

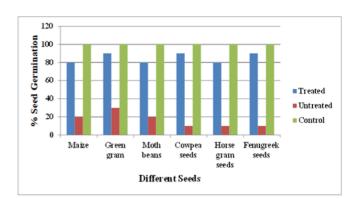
The results, summarized in Table 1, reveal significant improvements in industrial effluent samples over a period of 96 hours. The samples exhibited decreased turbidity and samples became odorless with neutral pH. Notably, a substantial 72% reduction in total solids was observed in the effluent samples. Furthermore, the bioremediation treatment was found to be effective in reducing heavy metal concentrations, with remarkable reductions observed for Zinc, Lead, Mercury, Copper, and Manganese (>85% reduction). These findings underscore the success of the bioremediation process in enhancing the quality of the industrial effluent by addressing both physicochemical parameters and heavy metal content. Detailed results are illustrated in Table 1

## Phytotoxic assay: Seed germination test

As depicted in Fig. 1, and Table 2, Green gram, Cowpea, and Fenugreek seeds exhibited a remarkable 90% germination rate, while Maize, Moth beans, and Horse gram seeds showed 80% germination rate in treated industrial effluent compared to control. In contrast, green gram seeds displayed 30% germination rate, while Maize and Moth beans exhibited 20% germination in untreated industrial effluent compared to the control. Cowpea, Horse gram, and Fenugreek seeds showed only 10% germination rate in untreated industrial effluent compared to control. (Fig, 1 and Table 2).

#### Phytotoxic assay: Pot .reulture test

The results of the pot culture test are represented in Table 3 & Table 4 and Fig. 2. It was observed that the percentage toxicity remarkably varied in treated roots (



**Figure 1.** Seed germination(%) by treated and untreated industrial effluent samples

		_ %				
Parameters	0	24 48		72	96	Reduction
Colour	Gray	Gray	Gray	Light Gray	Light Gray	
Odor	Fishy	Fishy	Fishy	Odourless	Odourless	
Temperature(°C)	29°C	28°C	28°C	28°C	28°C	
рН	0.00	3.03	5.4	7.70	08.84	
Turbidity (NTU)	0.00	23.56	41.24	66.60	79.14	79.14
TDS (mg/L)	0.00	22.06	47.97	63.50	71.18	71.18
TSS (mg/L)	0.00	27.54	43.37	61.05	72.91	72.91
TS (mg/L)	0.00	24.14	46.23	62.57	71.84	71.84
DO (mg/L)	1.36	1.92	2.89	4.15	4.89	
COD (mg/L)	0.00	22.30	52.01	68.45	79.78	79.78
BOD (mg/L)	0.00	25.37	54.25	73.10	80.63	80.63
Electrical Conductivity (mg/L)	0.00	10.72	28.40	52.75	62.61	62.61
Mercury (mg/L)	0.00	19.23	46.15	76.92	88.46	88.46
Lead (mg/L)	0.00	29.65	54.48	80.00	91.73	91.73
Zinc (mg/L)	0.00	23.43	59.37	83.85	95.31	95.31
Copper (mg/L)	0.00	25.47	58.49	76.41	86.79	86.79
Chromium (mg/L)	0.00	17.39	38.40	54.34	63.04	63.04
Cadmium mg/L)	0.00	14.28	41.75	60.43	72.53	72.53
Nickel (mg/L)	0.00	20.83	37.66	57.14	66.23	66.23
Iron(mg/L)	0.00	22.56	48.17	68.29	81.71	81.71
Manganese (mg/L)	0.00	26.08	43.47	60.86	88.41	88.41
Arsenic (mg/L)	0.00	14.77	32.95	50.00	64.77	64.77

## Table 1. Physicochemical, bioremediation and heavy metal analysis

Sr. No.	Name of the Seeds	Total	no of Seed	ls taken	Germinated Seeds			
		С	Т	U	С	Т	U	
1.	Maize	10	10	10	10	08	02	
2.	Green gram	10	10	10	10	09	03	
3.	Moth beans	10	10	10	10	08	02	
4.	Cowpea seeds	10	10	10	10	09	01	
5.	Horse gram seeds	10	10	10	10	08	01	
6.	Fenugreek seeds	10	10	10	10	09	04	
T = Treated, U= Untreated, C=Control								

## Table 2

Seed germination tests for treated and untreated industrial effluent samples

Sr. No	Control		Tre	eated	Untreated		
	Root	Shoot	Root	Shoot	Root	Shoot	
	Length	Length	Length	Length	Length	Length	
	(cm)	(cm)	(cm)	(cm)	(cm)	(cm)	
1	22.5	19.2	21.6	18.8	9.5	13.3	
2	24.5	20.1	23.5	19.7	10.0	10.5	
3	21.9	18.2	20.6	17.4	8.5	10.7	
4	23.1	21.4	21.1	18.8	9.0	14.6	
5	21.7	17.1	19.5	16.9	10.8	16.5	
6	21.3	18.2	20.6	17.2	8.7	10.8	
7	22.5	19.2	21.8	18.6	9.5	14.4	
8	23.8	18.1	21.8	17.8	13.5	14.8	
9	23.2	19.2	22.5	18.9	8.5	10.2	
10	20.5	16.4	19.1	16.1	10.5	13.1	

## Table 3

Pot culture test for treated and untreated industrial effluent samples

	Water sample					
Mean value	Control	Treated	Untreated			
Root	22.50	21.21	9.85			
Shoot	18.71	18.02	12.89			
Root: Shoot Ratio	1.20	1.17	0.76			
% Toxicity	0.00	2.50	36.66			

### Table 4

*Toxicity (%) of treated and untreated industrial effluents* 



A] Soaking of wheat



C] Growth of seedlings on 12th Day

B] Growth of seedlings on 5th Day



D] Measurement of root and shoot length

**Figure 2** Root and Shoot length during Pot culture test for treated industrial effluent sample

2.5%) and untreated roots (36.7%). Similarly, the root and shoot growth was numerically found to be higher in treated pots as compared to untreated

pots. One-way analysis of variance (ANOVA) for both, root (0.048) and shoot (0.003) was statistically significant (P<0.5) as shown in Table 5.

Table 5. ANOVA Test for pot culture study

Sr.				A	NOVA			
No.				(Single	factor-Root)			
1	Source of Variation		SS	df	MS	F	P-value	F crit
2	Between Groups		27.3045	9	3.033833	3.069128	0.047695	3.020383
3	Within Groups		9.885	10	0.9885			
		Total	37.1895	19				
Sr.				A	NOVA			
No.				(Single	factor-Shoot)			
1	Source of Variation		SS	df	MS	F	P-value	F crit
2	Between Groups		27.59	9	3.065611	6.529523	0.003545	3.02
3	Within Groups		4.7	10	0.4695	F	P-value	F crit
		Total	32.2855	19				

### **Discussions**

The comprehensive exploration of bio-remediation strategies employing metal-tolerant microbial isolates for the treatment of industrial effluents has yielded noteworthy results, as evidenced by the multifaceted assessment encompassing physicochemical parameters, heavy metal concentrations, and phytotoxicity assays.

## Physicochemical characteristics and bioremediation

The physicochemical analysis of the industrial effluent samples before and after the bioremediation process revealed substantial improvements. The observed reduction in turbidity, neutralization of pH, and the remarkable 72% decrease in total solids indicated the efficacy of the employed microbial isolates in enhancing the overall quality of the industrial effluent. These findings align with the anticipated outcomes of successful bio-remediation, where microbial activity contributes to the breakdown of organic and inorganic pollutants, leading to improved water clarity and reduced suspended solids. Moreover, the bioremediation treatment demonstrated its prowess in diminishing heavy metal concentrations in industrial effluent samples. Significant reductions of over 85% were noted for Zinc, Lead, Mercury, Copper, and Manganese. This outcome underscores the potential of the selected metal-tolerant bacterial strains (*Bacillus lichniformis, Pseudomonas aeruginosa, and Stenotrophomonas maltophilia*) in facilitating the removal or transformation of heavy metal pollutants, thereby ameliorating the environmental impact of industrial discharges.

## Phytotoxic assays

The seed germination test and pot culture test provided valuable insights into the phytotoxicity of the treated and untreated industrial effluents. Green gram, Cowpea, and Fenugreek seeds displayed a remarkable 90% germination rate in treated industrial effluent, emphasizing the positive impact of the bio-remediation process on seed viability. Conversely, untreated industrial effluent exhibited adverse effects on germination rates, with significantly lower percentages observed for various seeds. The pot culture study validated the positive influence of bio-remediation on plant growth. Treated roots demonstrated only 2.5% toxicity, in stark contrast to untreated roots with a greater toxicity rate (36.7%). This finding underscores the potential of metal-tolerant bacterial strains in mitigating the phytotoxic effects of industrial effluents, promoting healthier root development.

## Statistical Analysis:

Statistical analyses, including ANOVA, validated with the significance of observed differences in root and shoot growth between treated and untreated samples. The low p-values (p<0.5) reinforce the statistical robustness of the results, further supporting the positive impact of the bioremediation process on plant physiology.

### **Implications and Future Directions**

successful application of metal-tolerant The bacterial strains in the bio-remediation of industrial effluents holds promising implications for sustainable water management. The findings not only contribute to the understanding of microbial interactions with pollutants but also it could provide a practical framework for development of ecofriendly and efficient water treatment strategies. Future research avenues may explore the long-term stability and resilience of the introduced bacterial strains in real-world industrial effluent scenarios. Additionally, investigating the microbial community dynamics during bio-remediation and their interaction with different pollutants could further enhance the comprehension of the intricate processes involved.

## **Conclusion**

This study highlights the effectiveness of utilizing isolated heavy metal-tolerant bacterial strains for the bioremediation of industrial wastewater. This approach leads to improvements in physicochemical parameters and a reduction in heavy metal concentrations. The noticeable enhancement in germination rates and the improved growth of plants in the treated effluent underscore the promising potential for the sustainable application of these eco-friendly bacteria, making them a costeffective microbial bioremediation solution for industrial wastewater. Consequently, this research represents a significant advancement in the pursuit of sustainable solutions to alleviate the environmental impact of industrial activities. The incurporation of metal-tolerant microbial isolates into bioremediation processes emerges as a viable and effective strategy, striking a harmonious balance between industrial progress and environmental preservation.

## **References**

AYAZ T., KHAN S., KHAN A.Z., LEI M., ALAM M. (2020) Remediation of industrial wastewater using four hydrophyte species: A comparison of individual (pot experiments) and mix plants (constructed wetland). Journal of Environmental Management, 255:109833. https://doi.org/10.1016/j.jenvman.2019.109833

AZZAM A.M., TAWFIK A. (2015) Removal of heavy metals using bacterial bio-flocculants of Bacillus sp. and Pseudomonas sp. Journal of Environmental Engineering and Landscape Management, 23 (4):288–294. <u>https://doi.org/10.3846/16486897.2015.1068781</u>

HOBMAN J.L., BROWN N.L. (1996) Overexpression of MerT, the mercuric ion transport protein of transposon Tn501, and genetic selection of mercury hypersensitivity mutations. Molecular and General Genetics, 250 (1):129– 134. https://doi.org/10.1007/BF02191833

IANEVA O.D. (2009) Mechanisms of bacteria resistance to heavy metals. Mikrobiolohichnyi Zhurnal (Kiev, Ukraine), 71(6): 54–65. <u>https://pubmed.ncbi.nlm.nih.</u> gov/20455433/

IWASAKI Y., KAGAYA T., MIYAMOTO K., MATSUDA H. (2009) Effects of heavy metals on riverine benthic macroinvertebrate assemblages with reference to potential food availability for drift-feeding fishes. Environmental Toxicology and Chemistry, 28(2):354–363. https://doi.org/10.1897/08-200.1

JI G., SILVER S. (1955) Bacterial resistance mechanisms for heavy metals of environmental concern. Journal of Industrial Microbiology, 14(2):61–75. <u>https://doi.org/</u> 10.1007/BF01569887

MANISHA N, DINESH S, ARUN K. (2011) Isolation and characterisation of bacteria resistant to heavy metals Cadmium (Cd), Arsenic (As), Mercury (Hg) from industrial effluent. Global Journal of Applied Environmental Sciences, 1(2):127–132. <u>https://www.ripublication.com/</u> <u>gjaes/gjaesv1n202.pdf</u>

SAMADI M.T., SADEGHI S., RAHMANI A., SAGHI M.H. (2015) Survey of Water Quality in Moradbeik River Basis on WQI Index by GIS [Internet]. Rochester, NY. https://papers.ssrn.com/abstract=2610038

TAYANG A., SONGACHAN L.S. (2021) Microbial Bioremediation of Heavy Metals. *Current Science* 120(6):

1013. https://doi.org/10.18520/cs/v120/i6/1013-1025

TOLBATOV I., RE N., COLETTI C., MARRONE A. (2020) Determinants of the Lead(II) Affinity in pbrR Protein: A Computational Study. Inorganic Chemistry 59 (1):790–800. <u>https://doi.org/10.1021/acs.inorgchem.n 9b0 3059</u>

YUMPU.COM. (2023) Biosorption of Heavy Metals from Aqueous Solution using Bacillus Licheniformis. Avai lable from: <u>https://www.yumpu.com/en/document/</u> <u>view/53787901/biosorption-of-heavy-metals-from-</u> aqueous-solution-using-bacillus-licheniformis